

Glucosiduronates of 3 α ,21-Dihydroxy-5 β -pregnane-11,20-dione. Synthesis of C-3, C-21, and C-3,21 Derivatives¹

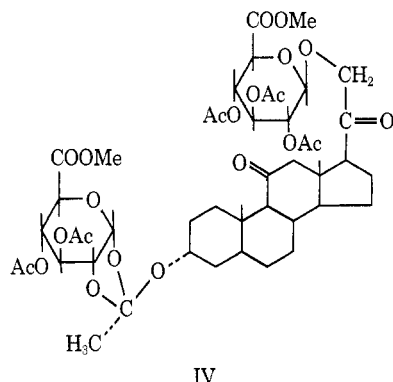
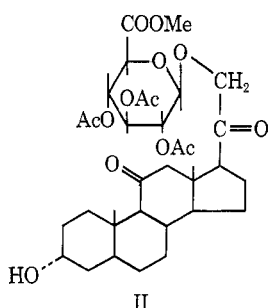
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On treatment with methyl 2,3,4-tri-*O*-acetyl-1-bromo-1-deoxy- α -D-glucuronate and Ag_2CO_3 , 3 α ,21-dihydroxy-5 β -pregnane-11,20-dione (I) yielded the corresponding steroidal 21-glucosiduronate (II), the 3,21-diglucosiduronate (III), the 21-glucosiduronate 3-glucuronosyl orthoacetate (IV), and the 3-glucosiduronate (V) as methyl ester acetates. In dilute methanolic HCl, IV was converted into II. Acetyl groups were removed from II, III, and V in methanol containing a catalytic amount of NaOH, and corresponding crystalline methyl esters VI, VII, and VIII were obtained. Alkaline hydrolysis of esters VI and VII followed by acidification gave the corresponding glucosiduronic acids. The 3-glucosiduronate (V) was converted into the C-20 semicarbazone (IX) to stabilize the ketolic group during alkaline hydrolysis; alkaline cleavage of the ester groups in IX followed by hydrolysis of the semicarbazone group at pH 2.1 gave 21-hydroxy-11,20-dioxo-5 β -pregnan-3 α -yl β -D-glucopyranosiduronic acid in 90% yield. Treatment of the 21-acetate of I with Ag_2CO_3 in benzene gave the 3-oxo derivative of I and showed that oxidation may occur as a side reaction in the Koenigs-Knorr procedure.

One of the principal pathways of metabolism of the adrenocortical hormones in man involves reduction of the 3-oxo- Δ^4 function to the 3 α -hydroxy-5 β -pregnane



structure. Subsequently, the 3 α -hydroxy metabolite is joined to glucuronic acid and the conjugate is excreted in the urine. Although 3 α -hydroxy-5 β -pregnanes which have either a ketolic side chain or a dihydroxyacetone function at C-17 are excreted principally as the C-3 glucosiduronic acids,^{2,3} small amounts of C-21 conjugates may also be present.^{4,5} 3 α ,21-Dihydroxy-5 β -pregnane-11,20-dione (1), a metabolite of corticosterone, occurs in human urine as a conjugate⁶ with glucuronic acid; however, the position(s) of attach-

ment of the conjugating group to the steroid has not been established unequivocally. In this paper we describe the synthesis of the C-3, the C-21, and the C-3,21 β -D-glucosyluronic acid derivatives of 1.

Treatment of the 3,21-dihydroxy compound 1 with 3 equiv of methyl 2,3,4-tri-*O*-acetyl-1-bromo-1-deoxy- α -D-glucuronate (hereafter referred to as methyl acetobromoglucuronate) under conditions reported previously⁷ gave a mixture which was fractionated by column chromatography to yield four substances: the C-21 glucosiduronate (4), the C-3,21 diglucosiduronate (12), the C-3 orthoacetate of the C-21 glucosiduronate (17), and the C-3 glucosiduronate (18).

The β configuration for the glucosidic linkage in compounds 4, 12, 17, 18, and 19 is suggested by the mode of synthesis,⁷ by the agreement of calculated and observed molecular rotations of the compounds (Table I), and by the hydrolysis of the glucosidic linkage of the

TABLE I
MOLECULAR ROTATIONS OF CONJUGATES

| Compd | Calcd, deg ^a | | Found, deg |
|-------|-------------------------|--------------------|------------|
| | α -Glycoside | β -Glycoside | |
| 4 | +950 | +244 | +159 |
| 5 | +1058 | +352 | +276 |
| 12 | +1555 | +143 | +98 |
| 18 | +950 | +244 | +283 |
| 19 | +1027 | +321 | +381 |

^a Values calculated as previously described.⁷

corresponding free glucosiduronic acids (7, 14, and 21) with β -glucuronidase. In addition, compounds 4 and 12 lack the nmr doublet ($\delta \cong 5.1$; $J_{1',2'} \cong 3.2$ Hz) which is characteristic⁸ of acetylated α -glycosides.

The C-21 glucosiduronate (4) was obtained from diol 1 in yields of 32–38%. This conjugate (4), as well as orthoacetate 17 which is derived from it, must have the glucosiduronate function at C-21 because it (4) does not reduce alkaline tetrazolium blue in the manner typical of α -ketolic steroids. However, compounds 4, 12, and 17 give a yellow color with this reagent, a response characteristic of the C-21 glucosiduronates of 11-dehydrocorticosterone and 11-deoxycorticosterone.⁹

(1) (a) This investigation was supported in part by Research Grant AM-5452 from the National Institutes of Health, Public Health Service. (b) Part of this investigation was presented at the 51st Annual Meeting of the Federation of American Societies for Experimental Biology, Chicago, Ill., April 16–21, 1967, Abstract No. 945.

(2) J. J. Schneider, M. L. Lewbart, P. Levitan, and S. Lieberman, *J. Amer. Chem. Soc.*, **77**, 4184 (1955).

(3) J. R. Pasqualini, *Bull. Soc. Chim. Biol.*, **45**, 277 (1963).

(4) J. R. Pasqualini and M.-F. Jayle, *C. R. Acad. Sci.*, **257**, 2345 (1963).

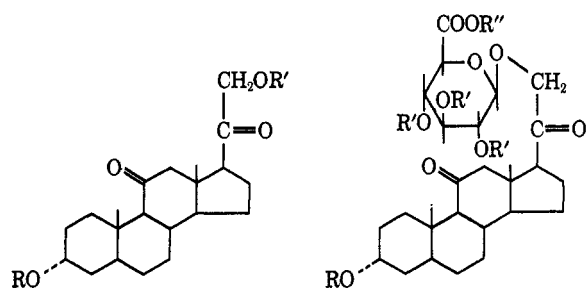
(5) R. Foggitt and A. E. Kellie, *Biochem. J.*, **91**, 209 (1964).

(6) J. R. Pasqualini, *J. Clin. Endocrinol. Metab.*, **27**, 885 (1967).

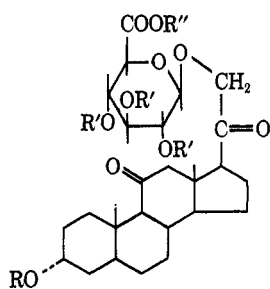
(7) V. R. Mattox, J. E. Goodrich, and W. D. Vrieze, *Biochemistry*, **8**, 1188 (1969).

(8) J. M. Van der Veen, *J. Org. Chem.*, **28**, 564 (1963).

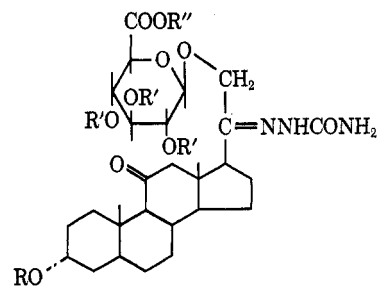
(9) V. R. Mattox, J. E. Goodrich, and W. D. Vrieze, *Steroids*, **18**, 147 (1971).



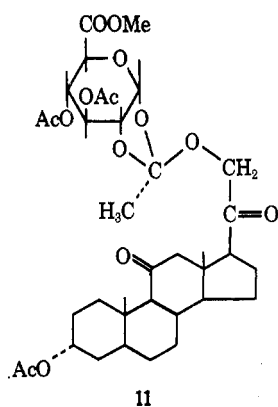
| | R | R' |
|---|----|----|
| 1 | H | H |
| 2 | Ac | H |
| 3 | H | Ac |



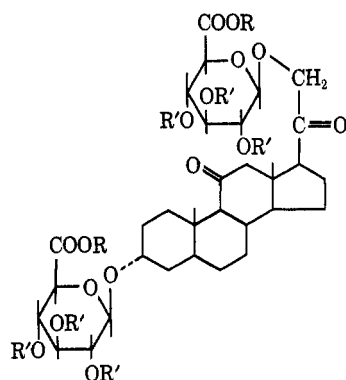
| | R | R' | R'' |
|---|----|----|-----------------|
| 4 | H | Ac | Me |
| 5 | Ac | Ac | Me |
| 6 | H | H | Me |
| 7 | H | H | H |
| 8 | H | H | NH ₄ |



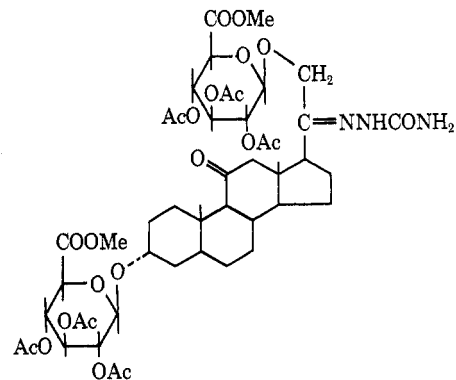
| | R | R' | R'' |
|----|----|----|-----|
| 9 | H | Ac | Me |
| 10 | Ac | Ac | Me |



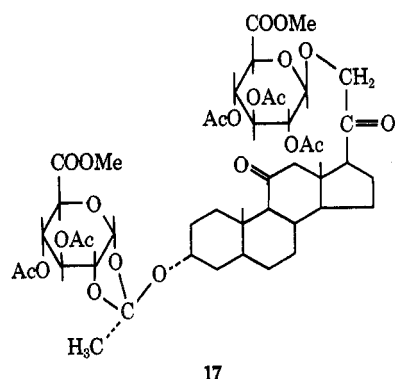
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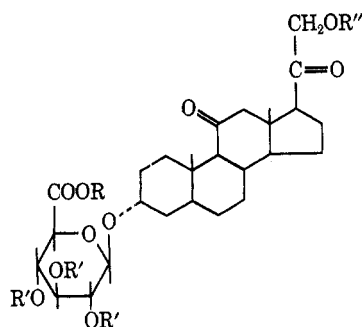
| | R | R' |
|----|-----------------|----|
| 12 | Me | Ac |
| 13 | Me | H |
| 14 | H | H |
| 15 | NH ₄ | H |



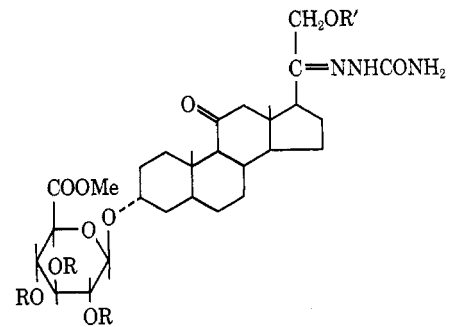
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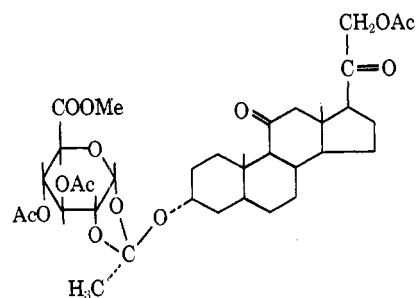
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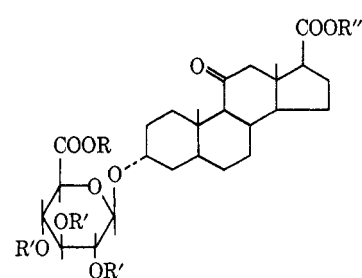
| | R | R' | R'' |
|----|----|----|-----|
| 18 | Me | Ac | H |
| 19 | Me | Ac | Ac |
| 20 | Me | H | H |
| 21 | H | H | H |



| | R | R' |
|----|----|----|
| 22 | Ac | H |
| 23 | Ac | Ac |
| 24 | H | H |



25



| | R | R' | R'' |
|----|----|----|-----|
| 26 | H | H | H |
| 27 | Me | Ac | H |
| 28 | Me | Ac | Me |

Conjugate **18**, obtained from **1** in 17% yield, has the glucosiduronate group at C-3; the substance reduces alkaline tetrazolium blue and is oxidizable by a catalytic amount of cupric acetate¹⁰ to a product which gives a positive reaction with the Porter-Silber reagent. In addition, acetylation of **18** gave the C-21 acetate (**19**) which could be prepared in 49% yield by treatment of 21-acetoxy-3 α -hydroxy-5 β -pregnane-11,20-dione (**3**) with methyl acetobromoglucuronate.

Efforts to crystallize the steroidal C-3,21 diglucosiduronate (**12**) after its preparation from diol **1** or from the C-21 monoglucosiduronate (**4**) were not successful; however, the amorphous diglucosiduronate (**12**) gave a crystalline C-20 semicarbazone (**16**) in 16–18% yield from **1**. The semicarbazone could be converted to crystalline 20-oxo diglucosiduronate (**12**) by treatment⁷ with pyruvic acid. Compounds **4**, **5**, **18**, **19**, and **20** also reacted with semicarbazide to give the corresponding semicarbazones which, on treatment with pyruvic acid, were converted to the 20-oxo starting materials.

The acetyl groups were removed from compounds **4**, **12**, **18**, and **19** by transesterification in methanol in the presence of dilute sodium methylate or sodium hydroxide, and the corresponding methyl esters were obtained in good yield. Removal of acetyl groups from a glucosiduronate moiety at C-3 occurs more slowly than from one at C-21.

Treatment of the C-21 glucosiduronate esters (**6** and **13**) with either ammonium hydroxide or sodium hydroxide followed by acidification gave the crystalline glucosiduronic acids (**7** and **14**). These acids were convertible back to the corresponding methyl esters (**6** and **13**) by treatment with diazomethane and to the parent dihydroxy steroid (**1**) by reaction with β -glucuronidase (Ketodase). The acetyl methyl ester glucosiduronates (**4** and **12**) were convertible to the corresponding free acids (**7** and **14**) in yields of about 80% without isolating the intermediate methyl esters.

When the acetylated glucosiduronate ester **19** was hydrolyzed with sodium hydroxide and the product was chromatographed on Celite in the presence of EDTA,¹⁰ glucosiduronic acid **21** was obtained in 47% yield along with some etianic acid (**26**) which was formed by alkaline cleavage¹¹ of the ketolic side chain. In the absence of EDTA, the copper which was eluted from the Celite caused oxidation of the ketolic group, and glucosiduronic acid **21** could not be crystallized. When sodium bicarbonate was used to hydrolyze ester **20**, glucosiduronic acid **21** was obtained in 66% yield.

The structure of etianic acid **26** was demonstrated by the following transformations. Reaction of the ketolic conjugate **18** with periodate yielded acid **27**, which, on treatment with diazomethane, gave ester **28**. Similarly, the etianic acid **26** was converted into methyl ester triacetate **28** by esterification with diazomethane followed by acetylation. In addition, the 17-carboxy steroidal conjugate (**27**) could be hydrolyzed with alkali to produce **26**.

Wendler, *et al.*,¹² showed indirectly that the α -ketolic side chain of a steroid is stable toward alkali

when this function is converted into a semicarbazone. We observed that, when semicarbazone **22** or **23** was treated with methanolic alkali, methyl ester semicarbazone **24** could be isolated in high yield. Reaction of this compound (**24**) with aqueous alkali followed by adjustment of the pH to 2.1 to hydrolyze the semicarbazone group gave crystalline glucosiduronic acid **21**. The yield of acid **21** from the acetate ester semicarbazone **23**, without isolation of the intermediates, was 88%. Removal of the semicarbazone function was also achieved by using the cation exchange resin, Dowex 50W-2X.

As a by-product of the Koenigs-Knorr reaction¹³ orthoacetate **17** was recovered in 4% yield when dihydroxy steroid **1** was treated with methyl acetobromoglucuronate. In the preparation of diglucosiduronate **12** from monoglucosiduronate **4**, orthoacetate **17** was obtained in 9% yield. In addition to the function at C-21, this substance has a glucosyluronate group attached to the steroid at C-3 through an orthoacetate structure which involves the acetyl group at C-2 of the carbohydrate, an α linkage to C-1 of the carbohydrate, and the oxygen of the 3 α -hydroxyl group of the steroid. The structure of orthoacetate **17** was suggested by its facile conversion to the C-21 glucosiduronate **4** during treatment with dilute acid,^{13c} and by the analytical values for C, H, CH₃O, and CH₃CO. The structure was confirmed by the nmr spectrum,^{13d,e} which has a band (δ 1.74 ppm; 3 protons) that is characteristic of the methyl group of an orthoacetate in the endo configuration relative to the carbohydrate group. This characteristic band was not present in either the C-21 glucosiduronate (**4**) or the C-3,21 diglucosiduronate (**12**).

In the preparation of the C-21 glucosiduronate (**5**) from the 21-hydroxy steroid (**2**), the corresponding steroid 21-yl glucuronylene orthoacetate (**11**) was obtained in 7% yield as a by-product. Similarly, orthoacetate **25** was obtained in 22% yield during preparation of **19**. These orthoacetates demonstrated characteristic spectral and chemical properties analogous to those of orthoacetate **17**.

Oxidation of a hydroxyl group to a ketone may occur in the Koenigs-Knorr reaction, as is shown by formation of 21-acetoxy-5 β -pregnane-3,11,20-trione from **3** during the preparation of **19**. This 3,11,20-trione also was formed, and isolated in a yield of 47%, by the action of silver carbonate on the 3-hydroxy compound **3** in boiling benzene in the absence of methyl acetobromoglucuronate. It was reported¹⁴ recently that primary and secondary alcohols are oxidized readily by silver carbonate on Celite in neutral media.

Experimental Section

Elemental analyses were carried out by Mr. Joseph F. Alicino, Metuchen, N. J.; samples were dried at 100° *in vacuo* immediately before analysis. Melting points were taken on a Fisher-Johns apparatus and are corrected. A substance synthesized by an optional procedure was identified by comparison of its melting point and ir spectrum with that of the authentic compound and by performing a mixture melting point determination. Infrared spectra were recorded with a Beckman IR-18 spectro-

(10) M. L. Lewbart and V. R. Mattox, *J. Org. Chem.*, **28**, 2001 (1963).

(11) L. Velluz, A. Petit, M. Pesez, and R. Barret, *Bull. Soc. Chim. Biol.*, **14**, 123 (1947).

(12) N. L. Wendler, Huang-Minlon, and M. Tishler, *J. Amer. Chem. Soc.*, **73**, 3818 (1951).

(13) (a) W. F. Goebel and F. H. Babers, *J. Biol. Chem.*, **110**, 707 (1935); (b) E. Pascu, *Advan. Carbohydr. Chem.*, **1**, 77 (1945); (c) H. R. Goldschmid and A. S. Perlin, *Can. J. Chem.*, **39**, 2025 (1961); (d) A. S. Perlin, *ibid.*, **41**, 399 (1963); (e) M. Mazurek and A. S. Perlin, *ibid.*, **43**, 1918 (1965).

(14) M. Fétizon and M. Golfier, *C. R. Acad. Sci., Ser. C*, **267**, 900 (1968).

photometer; nmr measurements were made with a Varian A-60 spectrometer with deuteriochloroform as solvent and tetramethylsilane as internal standard. Rotations were taken at $26^\circ \pm 1^\circ$ ($c \sim 1$). Evaporations were performed *in vacuo* on a rotary evaporator at a bath temperature of 40° . For column chromatography, Celite 545 was used as received from Johns-Manville and was impregnated with 40% of its weight of stationary (more dense) phase, using the solvent systems listed below. Holdback volume (HBV) represents the volume of the mobile phase retained by the packed portion of the column; the elution volume of a compound is expressed in HBV. Paper chromatography was performed as described previously;⁷ Zaffaroni technique was used with systems S1–S7 and Bush technique was used for systems S8–S14. Semicarbazones were detected by viewing chromatograms over 254-m μ radiation or by treating the chromatograms with 5% ethanolic phosphomolybdic acid; other compounds were detected by treating the chromatograms with alkaline tetrazolium blue.¹⁶ To detect acidic compounds which were not revealed by previously mentioned techniques, chromatograms were sprayed with a 0.04% solution of chlorophenol red in alcohol.¹⁶

Thin layer chromatography (tlc) was performed on silica gel G in 1:1 benzene-ether; compounds were detected by spraying the plates with water-sulfuric acid, 1:1, and charring.

Systems for Paper and Column Chromatography.—S1 = benzene-cyclohexane (25:75)-formamide-carbitol (1:1); S2 = benzene-cyclohexane (20:80)-formamide-carbitol (1:1); S3 = benzene-cyclohexane (25:75)-formamide; S4 = benzene-cyclohexane (50:50)-formamide; S5 = benzene-cyclohexane (75:25)-formamide; S6 = benzene-formamide; S7 = butyl acetate-formamide; mobile phase is butyl acetate saturated with formamide-water (1:1); S8 = cyclohexane-methanol-water (500:400:100); S9 = toluene-ethyl acetate-methanol-water (900:100:500:500); S10 = butyl acetate-butyl alcohol-water-acetic acid (90:10:90:10); S11 = butyl acetate-butyl alcohol-water-acetic acid (50:50:90:10); S12 = butyl acetate-butyl alcohol-water-acetic acid (25:75:90:10); S13 = butyl acetate-toluene-methanol-water-acetic acid (50:50:50:45:5); S14 = toluene-butyl acetate-methanol-water (25:75:50:50).

3 α ,21-Dihydroxy-5 β -pregnane-11,20-dione (1).—Treatment of 1.0-mg amounts of 7, 14, and 21 in separate flasks with 25,000 units of β -glucuronidase (Ketodase) under conditions previously described⁹ gave 1, which was identified by its chromatographic mobility on paper in systems S6, S7, and S9, and, after acetylation, in systems S1 and S8. For a previously prepared sample¹⁰ of 1, $[\alpha]_D^{25} = +99 \pm 2^\circ$ (CHCl₃). This value is used in the calculation of M_D for compound 4 (Table I).

3 α -Acetoxy-21-hydroxy-5 β -pregnane-11,20-dione (2). A.—To 8.65 g (20.0 mmol) of 3 α ,21-diacetoxy-5 β -pregnane-11,20-dione¹⁷ in 865 ml of methanol was added 8.65 g (86 mmol) of KHCO₃ in 288 ml of H₂O. After 1.5 hr the solution was acidified with acetic acid, concentrated, and extracted with chloroform. The extract was washed with NaHCO₃ solution and water and evaporated to dryness. The residue was chromatographed in system S1 on a column containing 700 g of Celite (HBV = 3.0), and crystals (4.52 g, mp 63–64°) were obtained from benzene. When a sample was dried at 1 mm and 100°, it melted and lost 15.3% (calcd for C₂₈H₄₆, 16.7%); $[\alpha]_D^{25}$ on the dried sample = $+116 \pm 2^\circ$ (CHCl₃). A sample, dried to constant weight at 60° and 1 mm, lost 12.6% (calcd for loss of $\frac{3}{4}$ C₆H₆, 12.5%), and absorbance of the dried product at 255 m μ indicated 4.4% benzene (calcd for $\frac{1}{4}$ C₆H₆, 4.8%): ir (KBr) 3450 (OH), 1730 (acetate C=O), 1702 (ketone C=O), and 1237 cm⁻¹ (ester COC). *Anal.* Calcd for C₂₈H₄₄O₅· $\frac{1}{4}$ C₆H₆: C, 71.76; H, 8.73. Found: C, 71.65; H, 8.74.

B. From 11.—To 100 mg of orthoacetate 11 in 10 ml of benzene was added 1.0 ml of 0.1 N HCl in methanol.¹⁸ After 10 min, the solution was washed with 1.0 N Na₂CO₃ and then with water until neutral and taken to dryness. The residue was chromatographed on 30 g of Celite in system S1 (HBV 3.6). Crystals (18 mg, 28%, mp 63–65°) were obtained from benzene-cyclohexane and identified as 2.

21-Acetoxy-3 α -hydroxy-5 β -pregnane-11,20-dione (3). A. **From 1.**—A pyridinium acetate buffer was prepared by diluting 121 ml of pyridine to 500 ml with glacial acetic acid. To 14.0 g

of compound 1¹⁰ was added 500 ml of glacial acetic acid and 500 ml of pyridinium acetate buffer at 25°. While the flask was being shaken, 100 ml of acetic anhydride was added; the mixture became homogeneous in 15 min. One hour later the mixture was worked up, and crystals (8.2 g, 52%, mp 138–141°) were obtained from ethanol. In a similar preparation in which the product was chromatographed in system S5, 3 emerged at 2.0 HBV (yield 65%, mp 142–143°), $[\alpha]_D^{25} +108 \pm 2^\circ$ (CHCl₃) [reported¹⁷ mp 137–138°; $[\alpha]_D^{25} 109^\circ$ (CHCl₃)].

B. From 25.—To 100 mg of orthoacetate 25 in 10 ml of benzene was added 1.0 ml of 0.1 N HCl in methanol. After 10 min the solution was washed twice with 1.0 N Na₂CO₃ and then with water until neutral and taken to dryness. Crystalline 3 (83%, mp 141–143°) was obtained from benzene-cyclohexane.

Methyl (3 α -Hydroxy-11,20-dioxo-5 β -pregnan-21-yl 2,3,4-Tri-O-acetyl- β -D-glucopyranosid)uronate (4). A. **From 1.**—A mixture of 348 mg (1.0 mmol) of 1 and 1.1 g (4.0 mmol) of Ag₂CO₃ in 150 ml of benzene was treated with 1.13 g (3.0 mmol) of methyl 2,3,4-tri-O-acetyl-1-bromo-1-deoxy- α -D-glucuronate by the general procedure described previously.⁷ The product was chromatographed in solvent system S4; 9.2-ml fractions were collected. To monitor the effluent, alternate fractions were mixed with 1 ml of water and 50- μ l samples of organic phase were removed, taken to dryness, and mixed with 1.0 ml of concentrated H₂SO₄. After 2 hr, the absorbance was read at 415 m μ and plotted against fraction number. Additional samples (25 μ l) were removed, spotted on filter paper, and treated with alkaline tetrazolium blue. For fractions 23–37 (HBV 1.8) see 17 from 1; for fractions 38–53 (HBV 2.6) see 16 from 1 *via* 12; for fractions 69–94 (HBV 4.6), see 18 from 1.

Fractions 189–259 (HBV 13.5) were combined, washed with water, and concentrated. Crystals of glucosiduronate 4 (215 mg, 32%) were obtained from ethanol: mp 108–111°; homogeneous in system S5; $[\alpha]_D^{25} +24 \pm 2^\circ$ (CHCl₃); ir (KBr) 3520 (OH), 1757 (ester C=O), 1705 (ketone C=O), and 1235 sh, 1215 cm⁻¹ (ester COC); nmr (CDCl₃) δ 0.57 (C-18 CH₃), 1.15 (C-19 CH₃), 1.81 (C-3 OH?), 2.01 (two OAc), 2.08 (one OAc), 2.45 (C-12 protons), 3.75 (CH₂O of ester), and 4.50 ppm (C-21 CH₂O). *Anal.* Calcd for C₃₄H₄₈O₁₃: C, 61.43; H, 7.28. Found: C, 61.61; H, 7.29.

B. From 9.—Treatment of semicarbazone 9 (200 mg) in chloroform with pyruvic acid⁷ gave crystals of 4, mp 105–108°, in 79% yield.

C. From 17.—Orthoacetate 17 (30 mg) was treated as described for the preparation of 2 from 11; crystals (4.9 mg, 24% yield, mp 103–105°) were obtained from cold ethanol and identified as glucosiduronate 4.

Methyl (3 α -Acetoxy-11,20-dioxo-5 β -pregnan-21-yl 2,3,4-Tri-O-acetyl- β -D-glucopyranosid)uronate (5). A. **From 2.**—A solution of 2 (3.38 g) in benzene was treated with methyl 2,3,4-tri-O-acetyl-1-bromo-1-deoxy- α -D-glucuronate by the procedure described for the preparation of 4 from 1, and the product was chromatographed on 700 g of Celite in system S1. The band eluted at 2.0 HBV contained orthoacetate 11 (see 11 from 2). The band eluted at 3.0 HBV gave crystals of 5 from benzene-cyclohexane (1.62 g, 27%, mp 168–169°, homogeneous by tlc).

B. From 4.—Treatment of 200 mg of 4 with a mixture of 1.0 ml each of acetic anhydride and pyridine at room temperature for 3 hr followed by crystallization of the product from methanol gave homogeneous (system S5) conjugate 5 (162 mg, 76%, mp 169–170°). The pure substance had mp 170–171°; $[\alpha]_D^{25} +39 \pm 2^\circ$ (CHCl₃); ir (KBr) 1775, 1750 (acetate CO), 1725 (ester CO), 1702 (ketone C=O), and 1245, 1215 cm⁻¹ (ester COC). *Anal.* Calcd for C₃₈H₅₀O₁₄· $\frac{1}{2}$ H₂O: C, 60.40; H, 7.32; CH₃CO, 24.06. Found: C, 60.53; H, 6.94; CH₃CO, 24.78.

Compound 5 was prepared also by acetylation of 6 and by treatment⁷ of 10 with pyruvic acid.

Methyl (3 α -Hydroxy-11,20-dioxo-5 β -pregnan-21-yl β -D-Glucopyranosid)uronate (6). A. **From 4.**—To a solution of 665 mg of triacetate 4 in 40 ml of dry methanol was added 0.30 ml of 1.2 N sodium methylate in methanol. After 30 min a slight excess of acetic acid was added, the solution was evaporated to dryness, and crystals (393 mg, 72%, mp 203–205°; homogeneous in system S13) were obtained from cold ethanol: $[\alpha]_D^{25} +37 \pm 2^\circ$ (CH₂OH); ir (KBr) 3420, 3300 (OH), 1746 (ester C=O), and 1708 cm⁻¹ (C-11 + C-20, C=O). *Anal.* Calcd for C₂₈H₄₂O₁₀· $\frac{1}{2}$ H₂O: C, 61.41; H, 7.91; CH₃O, 5.66. Found: C, 61.78; H, 7.50; CH₃O, 6.03.

B. From 7.—Treatment of acid 7 with diazomethane gave an ester (mp 202–204°) which was identical with 6.

(15) R. Neher, "Steroid Chromatography," 2nd ed, Elsevier, New York, N. Y., 1964, p 122.

(16) M. L. Lewbart and V. R. Mattox, *J. Org. Chem.*, **28**, 1779 (1963).

(17) R. Deghenghi and C. R. Engel, *J. Amer. Chem. Soc.*, **82**, 3201 (1960).

3 α -Hydroxy-11,20-dioxo-5 β -pregnan-21-yl β -D-Glucopyranosiduronic Acid (7). A. From 4.—To a solution of 382 mg of triacetate 4 in 7.5 ml of chloroform and 3.75 ml of methanol was added 3.75 ml of 0.04 *N* NaOH in methanol. The mixture stood at room temperature for 45 min; then 15 ml of methanol and 7.0 ml of 1.0 *N* aqueous NaOH were added. The mixture stood at room temperature for an additional 30 min, the pH was brought to 4 with dilute H₂SO₄, and the precipitate of sodium sulfate was filtered off and washed with 10 ml of methanol; the filtrate was concentrated to about 5 ml to remove the chloroform and methanol. Water was added to a volume of 80 ml, the pH was brought to 2.1 with H₂SO₄, and the homogeneous mixture was poured onto a column containing 40 g of Amberlite XAD-2.^{7,18,19} The column was washed twice with 80 ml of water and three times with 80 ml of ethanol. The ethanol eluates contained an impurity which was removed by chromatography in system S10. A band which emerged at 2.1 HBV gave crystals (241 mg, 79%) from methanol-ethyl acetate (mp 155–157.5°) which were identical with 7 prepared from 8.

B. From 8.—A solution of ammonium salt 8 (300 mg) in 30 ml of water was adjusted to pH 3.0 with H₂SO₄ and concentrated almost to dryness. The (NH₄)₂SO₄ was separated by its insolubility in dry butyl alcohol, and 7 was crystallized from water and then from methanol-ethyl acetate (yield 82%, mp 157–160°): homogeneous in S10; [α]_D +35 \pm 2° (CH₃OH); ir (KBr) 3410 (OH), 1700 cm⁻¹ (C=O). *Anal.* Calcd for C₂₇H₄₀O₁₀·1/2H₂O: C, 60.77; H, 7.75. Found: C, 60.73; H, 7.97.

Ammonium (3 α -Hydroxy-11,20-dioxo-5 β -pregnan-21-yl β -D-Glucopyranosid)uronate (8). A. From 6.—A solution of methyl ester 6 (1.0 mmol) in 50 ml of 1 *N* NH₄OH stood at room temperature for 30 min and was evaporated to dryness. Crystals were obtained (82%, mp 167–170° dec) from methanol-ethyl acetate: homogeneous in system S10; [α]_D +35 \pm 2° (CH₃OH); ir (KBr) 3400–3220 (OH and NH), 1703 (C-11 + C-20, C=O), 1600 (carboxyl C=O), and 1405 cm⁻¹ (COO⁻). *Anal.* Calcd for C₂₇H₄₀O₁₀N: C, 59.87; H, 8.00. Found: C, 59.40; H, 7.60.

Compound 8 was also prepared from acid 7.

Methyl (3 α -Hydroxy-11,20-dioxo-5 β -pregnan-21-yl 2,3,4-Tri-O-acetyl- β -D-glucopyranosid)uronate 20-Semicarbazone (9).—A solution of compound 4 (200 mg) was treated⁷ with semicarbazide hydrochloride to give crystals of 9 (homogeneous in system S6; 91% yield, mp 152–153°): uv max (CH₃OH) 236 m μ (ϵ 12,300); ir (KBr) 3590, 3495, 3350 (OH and NH), 1758 (ester C=O), 1700 (C-11 C=O), 1690 sh (amide C=O), 1562 (amide), and 1225 sh, 1207 cm⁻¹ (ester COC). *Anal.* Calcd for C₃₅H₅₁O₁₃N₃: N, 5.82. Found: N, 5.65.

Methyl (3 α -Acetoxy-11,20-dioxo-5 β -pregnan-21-yl 2,3,4-Tri-O-acetyl- β -D-glucopyranosid)uronate 20-Semicarbazone (10).—10 was obtained in 94% yield from 5: hygroscopic crystals; mp 165–167° from methanol; homogeneous in S6; uv max (CH₃OH) 236 m μ (ϵ 13,000); ir (KBr) 3490, 3360 (NH), 1757 (acetate C=O), 1735 sh (ester C=O), 1702 (C-11 C=O), 1690 sh (amide C=O), 1570 (amide), and 1235, 1212 cm⁻¹ (ester COC). *Anal.* Calcd for C₃₇H₅₃O₁₄N₃·1/2H₂O: C, 57.50; H, 7.01; N, 5.44. Found: C, 57.25; H, 6.61; N, 5.21.

Methyl α -D-Glucopyranosidurate Cyclic 1,2-(Hydrogen [8]-Orthoacetate) 3,4-Diacetate 21-Ester with 3 α ,21-Dihydroxy-5 β -pregnane-11,20-dione 3-Acetate (11).—Fractions 93–144 (HBV 2.0), described under 5 from 2, were combined, washed with water, and taken to dryness. The residue was chromatographed on 300 g of Celite in system S2 (HBV 2.8) and crystals of 11 (385 mg, 6.7%) were obtained from methanol-water: homogeneous by tlc and in system S1; mp 91–94°; [α]_D +68 \pm 2° (CHCl₃); ir (KBr) 1753 (acetate C=O), 1735 sh (ester C=O), 1705 (C-11 + C-20, C=O), and 1232, 1208 sh cm⁻¹ (ester COC); nmr (CDCl₃) δ 0.58 (C-18 methyl), 1.17 (C-19 methyl), 1.75 (orthoacetate methyl, endo), 2.02 (C-3 OAc), 2.08 and 2.12 (C-4' and C-5' OAc), 2.47 (C-12), 3.02 [C-17 (?)], 3.78 (CH₂O of ester), 3.97 (C-21 methylene), 4.75 (C-3 proton), and 5.85 ppm (*J* = 5 Hz) (C-1' proton). *Anal.* Calcd for C₃₈H₅₆O₁₄: C,

61.17; H, 7.13; CH₃O, 4.39; CH₃CO, 24.36. Found: C, 60.98; H, 7.31; CH₃O, 4.35; CH₃CO, 22.53.

11,20-Dioxo-5 β -pregnan-3 α ,21-ylene Di(methyl 2,3,4-Tri-O-acetyl- β -D-glucopyranosiduronate) (12). A. From 13.—Acetylation of diglucosiduronate dimethyl ester 13 with 1:1 acetic anhydride-pyridine gave a product identical with ester 12 below.

B. From 16.—Semicarbazone 16 (2.08 g) was treated⁷ with pyruvic acid as described previously to give 12, which was crystallized from a mixture of ethanol-acetone-water (1.77 g, 90%, mp 125–128°): homogeneous in system S5 and by tlc; [α]_D +10 \pm 2° (CHCl₃); ir (KBr) 1753 (ester C=O), 1706 (C-11 + C-20 C=O), and 1230 sh, 1210 cm⁻¹ (ester COC). *Anal.* Calcd for C₄₇H₆₄O₂₂: C, 57.54; H, 6.58; CH₃CO, 26.32. Found: C, 57.19; H, 6.46; CH₃CO, 24.64.

Dimethyl 11,20-Dioxo-5 β -pregnan-3 α ,21-ylene Di- β -D-glucopyranosiduronate (13). A. From 12.—Ester 13 was prepared from 12 in 86% yield as described for ester 6 from triacetate ester 4. Crystals were obtained from a mixture of methanol and *tert*-butyl alcohol (mp 147–150°): homogeneous in S10; [α]_D +17 \pm 2° (CH₃OH); ir (KBr) 3410 (OH), 1742 (ester C=O), and 1702 cm⁻¹ (C-11 + C-20 C=O). *Anal.* Calcd for C₃₅H₅₃O₁₆: C, 57.68; H, 7.19. Found: C, 57.50; H, 7.61.

B. From 14.—Treatment of acid 14 with diazomethane also gave ester 13.

11,20-Dioxo-5 β -pregnan-3 α ,21-ylene Di(β -D-glucopyranosiduronic Acid) (14). A. From 12.—Compound 14 was prepared from diglucosiduronate 12 (0.25 mmol) as described for preparation of 7 from 4. The residue from the Amberlite XAD-2 column contained material (chromatography system S12) which gave an atypical color (purple) with alkaline tetrazolium blue (compound 14 gave a yellow color). The residue was chromatographed on 50 g of Celite in system S11 (HBV 6.6) and the conjugate was crystallized from methanol-ethyl acetate (144 mg, 83%, mp 185° dec) and identified as 14.

B. From 15.—A solution of ammonium salt 15 (200 mg) in water was acidified to pH 3 with H₂SO₄ and taken almost to dryness. The residue was extracted with two 100-ml portions of absolute ethanol, the solution was taken to dryness, and crystals of 14 were obtained from methanol-ethyl acetate (156 mg, 82%, mp 175° dec): homogeneous in system S11; [α]_D +17 \pm 2° (CH₃OH); ir (KBr) 3430 (OH), 1720 sh (carboxyl C=O), and 1703 cm⁻¹ (C-11 + C-20 C=O). *Anal.* Calcd for C₃₅H₄₈O₁₆·1/2H₂O: C, 55.84; H, 6.96. Found: C, 55.77; H, 6.76.

C. From 16.—The diglucosiduronate semicarbazone 16 (1.038 g) was converted into acid 14 by the procedure used to prepare 7 from 4, except for the following modification. The solution stood at pH 2.1 for 1 hr (in order to hydrolyze the semicarbazone) before it was poured into the column containing Amberlite XAD-2; the column was then washed with two 80-ml portions of 0.01 *N* H₂SO₄ before being washed with three 80-ml portions of water, etc. Crystals of 14 (529 mg, 75%, mp 170–174°) were obtained from methanol-ethyl acetate.

Diammonium 11,20-Dioxo-5 β -pregnan-3 α ,21-ylene Di- β -D-glucopyranosiduronate (15).—A solution of ester 13 (200 mg) in 20 ml of 1 *N* NH₄OH stood for 2 hr at 25° and was concentrated to dryness. Crystals (188 mg, 93%) of 15 were obtained from methanol-ethanol: mp 178–180° dec; homogeneous in system S11; [α]_D +10 \pm 2° (CH₃OH); ir (KBr) 3400–3220 (OH and NH), 1700 (C-11 + C-20 C=O), 1600 (carboxyl C=O), and 1400 cm⁻¹ (COO⁻). *Anal.* Calcd for C₃₈H₅₄O₁₆N₂·H₂O: C, 52.65; H, 7.44. Found: C, 52.66; H, 6.75.

11,20-Dioxo-5 β -pregnan-3 α ,21-ylene Di(methyl 2,3,4-Tri-O-acetyl- β -D-glucopyranosiduronate) 20-Semicarbazone (16). A. From 1 via 12.—Fractions 38–53 (HBV 2.6), described under the preparation of 4 from 1, were combined, washed with water, and taken to dryness. The product (12) would not crystallize as the ketone; it was converted⁷ into semicarbazone 16 (130 mg, 13% yield from 1, mp 174–175°), a homogeneous (system S6), hygroscopic product. When recrystallized from methanol, it had mp 179–180°; uv max (CH₃OH) 236 m μ (ϵ 12,800); ir (KBr) 3510, 3380 (NH), 1757 (ester C=O), 1702 (C-11 + amide, C=O), 1570 (amide), and 1230 sh, 1212 cm⁻¹ (ester COC). *Anal.* Calcd for C₄₈H₆₇O₂₂N₃·1/2H₂O: C, 55.06; H, 6.55; N, 4.02; CH₃CO, 24.67. Found: C, 54.90; H, 6.23; N, 3.78; CH₃CO, 21.83.

B. From 4 via 12.—Fractions corresponding to 2.6 HBV (described under preparation of 17 from 4) were combined and converted into 16 (254 mg, 24% from 4, mp 176–177°) as described above.

(18) This resin is very useful for separating steroidal glucosiduronic acids from inorganic acids, salts, and various other water-soluble compounds. We have employed it to separate steroidal glucosiduronic acids from ammonium sulfate, sodium sulfate, ammonium chloride, semicarbazide (after acid hydrolysis of a semicarbazone), inorganic acids in general, and debris from the effluent of a Celite column when the solvent system contained aqueous acetic acid. Also, we have used it for converting ammonium, sodium, potassium, thallium, and barium glucosiduronates into free acids.

(19) H. L. Bradlow, *Steroids*, **11**, 265 (1968).

C. From 12.—Treatment⁷ of 12 (490.5 mg) with semicarbazide hydrochloride yielded a product (512 mg, 99%, mp 177–179°) identical with authentic 16.

Methyl 3 α -Hydroxy-11,20-dioxo-5 β -pregnan-21-yl β -D-Glucopyranosiduronate 2,3,4-Triacetate 3'-(Dihydrogen Orthoacetate) Cyclic 1,2-(S)-Ester with Methyl α -D-Glucopyranuronate 3,4-Diacetate (17). A. From 1.—Fractions 23–37 (HBV 1.8), obtained during preparation of 4 from 1, gave a residue which was chromatographed on 100 g of Celite in solvent system S3. The appropriate fraction (HBV 13.5) gave crystals (43 mg, 4.4%, mp 110–115°) of orthoacetate 25 from methanol-water; this product was identical with orthoacetate 25 described in the following paragraph.

B. From 4.—One millimole of 3 α -hydroxy conjugate 4 was treated with 3 mmol of methyl acetobromoglucuronate as described for the preparation of 4 from 1. The product was chromatographed on 100 g of Celite (system S4). (For the substance eluted at 2.6 HBV, see 16 from 12, B.) Fractions corresponding to 1.8 HBV were combined, washed with water, and taken to dryness. The residue was rechromatographed in solvent system S3 on 100 g of Celite. The residue from the principal band (HBV 16) weighed 127 mg, a 13% yield of orthoacetate 17. Crystals (90 mg, 9.2%, mp 111–116°) from benzene-cyclohexane were homogeneous by tlc; ir (KBr) 1758 (ester C=O), 1705 (C-11 + C-20 C=O), and 1212 cm⁻¹ (ester COC); nmr (CDCl₃) δ 0.55 (C-18 CH₃), 1.13 (C-19 CH₃), 1.74 (orthoacetate CH₃), 2.01 (two OAc groups of C-21 glucuronyl group), 2.09 (one OAc group of C-21 glucuronyl group; two OAc groups of C-3 glucuronyl function), 2.43 (probably C-12 protons), 3.75 (CH₃O of ester), and 4.50 ppm (C-21 methylene). *Anal.* Calcd for C₄₇H₆₄O₂₂·H₂O: C, 56.50; H, 6.66; CH₃O, 6.21; CH₃CO, 25.85. Found: C, 56.40; H, 6.45; CH₃O, 7.05; CH₃CO, 24.78.

Methyl (21-Hydroxy-11,20-dioxo-5 β -pregnan-3 α -yl 2,3,4-Tri-O-acetyl- β -D-glucosid)uronate (18). A. From 1.—Fractions 69–94 (HBV 4.6), described under 4 from 1, were combined, washed with water, and freed of solvent. Crystals (113 mg, 17%, mp 113–116°), obtained from acetone-methanol-water, were homogeneous in system S5; [α]_D +42 \pm 2° (CHCl₃); ir (KBr) 3490 (OH), 1755 (ester C=O), 1702 (C-11 + C-20 C=O), and 1210 cm⁻¹ (ester COC). *Anal.* Calcd for C₃₄H₄₈O₁₃·1/2H₂O: C, 60.60; H, 7.33. Found: C, 60.68; H, 7.07.

Compound 18 reduced alkaline tetrazolium blue and reacted with cupric acetate¹⁰ to produce a product which gave a yellow color with Porter-Silber reagent.

B. From 22.—Treatment of semicarbazone 22 (20 mg) with pyruvic acid⁷ produced 9 mg (49%, mp 110–113°) of 18.

Methyl (21-Acetoxy-11,20-dioxo-5 β -pregnan-3 α -yl 2,3,4-Tri-O-acetyl- β -D-glucosid)uronate (19). A. From 3.—Compound 3 (3.9 g, 10.0 mmol) was treated with methyl 2,3,4-tri-O-acetyl-1-bromo-1-deoxy- α -D-glucuronate as described under the preparation of 4 and the product was chromatographed in system S4. Fractions were combined to give residues designated as follows: 1.5 HBV (a mixture of glucosiduronate 19 and orthoacetate 25; see below in this paragraph); 2.2 HBV (the 3-oxo compound, 21-acetoxy-5 β -pregnane-3,11,20-trione; see below). The residue designated 1.5 HBV gave crystals (3.02 g, 43% mp 106–109°) of 19 from benzene-cyclohexane: homogeneous in system S1; [α]_D +54 \pm 2° (CHCl₃); nmr (CDCl₃) δ 0.60 (C-18 CH₃), 1.12 (C-19 CH₃), 2.02 (2',3',4' OAc groups), 2.16 (C-21 OAc), 2.50 (C-12 protons), 3.5 (CH₃O of ester), and 4.55 ppm (C-21 methylene). The ir spectrum was identical with that of compound 19. (For isolation of the 3-orthoacetate 25 from the mother liquor of 19, see 25 below.)

B. From 18.—Acetylation of 18 (200 mg) followed by crystallization from methanol-water gave 19 (174 mg, 81%, mp 106–108°): [α]_D +54 \pm 2° (CHCl₃); ir (KBr) 1757 (ester C=O), 1728 sh (C-20 C=O), 1707 (C-11 C=O), and 1210 cm⁻¹ (ester COC). The sample for analysis was dried *in vacuo* at 78°. *Anal.* Calcd for C₃₈H₅₀O₁₄: C, 61.17; H, 7.13; CH₃CO, 24.36. Found: C, 60.99; H, 7.05; CH₃CO, 23.31.

Compound 19 was prepared from 20 in a similar manner and from 23 by treatment with pyruvic acid.

Methyl (21-Hydroxy-11,20-dioxo-5 β -pregnan-3 α -yl β -D-Glucopyranosid)uronate (20). A. From 19.—Treatment of 353 mg of compound 19 with 0.01 N NaOH in 15 ml of chloroform-methanol, 1:1, for 45 min followed by addition of an excess of acetic acid and 1.0 mg of EDTA, concentration *in vacuo*, and separation from ethyl acetate-cyclohexane yielded amorphous 20 which was homogeneous in system S13 (232 mg, 86%):

[α]_D +51 \pm 2° (CH₃OH); ir (KBr) 3440 (OH), 1746 (ester C=O), and 1705 cm⁻¹ (C-11 + C-20 C=O). *Anal.* Calcd for C₂₈H₄₂O₁₀·H₂O: C, 60.41; H, 7.97; CH₃O, 5.57. Found: C, 60.58; H, 7.77; CH₃O, 5.62.

B. From 21.—Esterification of acid 21 with diazomethane also gave ester 20.

21-Hydroxy-11,20-dioxo-5 β -pregnan-3 α -yl β -D-Glucopyranosiduronic Acid (21). A. From 19.—Compound 19 was treated as described below under procedure 1 for preparation of 21 from 23. It was not possible to obtain crystals. The product was freed of salt and inorganic acid by use⁷ of Amberlite XAD-2; to the alcoholic eluate was added 2 mg of EDTA, the solution was taken to dryness, and the residue was chromatographed on 100 g of Celite (system S10). Bands emerged at 1.14 (see 26 from 19) and at 1.8 HBV. EDTA (2 mg) was added to the eluate containing the latter band, the solvent was removed, and crystals of 21 were obtained from 0.01 N H₂SO₄ and washed with H₂O (122 mg, 47%, mp 156–158°).

In two similar experiments on the preparation of 21 from 19 in which only 0.10 mg of EDTA was added to the alcoholic eluate from the Amberlite column and again after Celite chromatography, crystals could not be obtained. Several solvents were used during a period of 2 days. A paper chromatogram (system S10) showed that the original ketolic compound 21 (*R*_f 0.29) had disappeared and that a new Porter-Silber-positive¹⁰ compound (*R*_f 0.70; presumably the 20-keto-21-aldehyde) had been formed.

In another experiment, the mobile phase was run through a column of Celite like that used for chromatography of 21; the fraction corresponding to the band at 1.8 HBV was collected, taken to dryness, and shown to contain (by atomic absorption) 1.5 μ g of copper. To one-tenth of this residue was added 17 mg of compound 21 in 0.10 ml of methanol; after 24 hr, the solution gave a strong color with the Porter-Silber reagent. Color with the Porter-Silber reagent was not obtained when 17 mg of compound 21 was treated similarly with 0.10 mg of the residue in the presence of 0.10 mg of EDTA.

B. From 23. Procedure 1.—Semicarbazone 23 (764 mg) was treated with alkali as described under 7 from 4. Methanol²⁰ (10.0 ml) was added to dissolve the oil which separated when the pH was adjusted to 2.1. After 1 hr, the solution was concentrated *in vacuo* to 7 ml. Crystals, which formed slowly, were filtered, washed well with cold water, and dried at 100° (497 mg, 93%, mp 157–159°). In a second preparation the yield was 88% (mp 158–160°): homogeneous in system S10; [α]_D +59 \pm 2° (CH₃OH); ir (KBr) 3400 (OH), 1748 sh (carboxyl C=O), and 1700 cm⁻¹ (C-11 + C-20 C=O). *Anal.* Calcd for C₂₇H₄₀O₁₀·1/2H₂O: C, 60.77; H, 7.75. Found: C, 60.76; H, 7.72.

B. From 23. Procedure 2. Ion-Exchange Column.—One millimole of 23 was treated with alkali as described in the previous paragraph and the solution was acidified with acetic acid, concentrated to dryness, and processed⁷ on a column of Dowex 50W-X2 (40 g); acid 21 was obtained in 82% yield.

C. From 20.—A solution which contained 100 mg of 20 in 20 ml of 0.1 M NaHCO₃ stood at 26° for 24 hr. The pH was adjusted to 2.1 with 0.5 M H₂SO₄, and the solution was concentrated and cooled. Crystals of 21 (mp 157–159°) were obtained in 40% yield. By desalting the filtrate on Amberlite XAD-2 and chromatographing the steroid residue in system S10, an additional 26% of 21 was obtained.

Methyl (21-Hydroxy-11,20-dioxo-5 β -pregnan-3 α -yl 2,3,4-Tri-O-acetyl- β -D-glucosid)uronate 20-Semicarbazone (22).—22 was obtained from 18 by the procedure described in the next paragraph and crystallized from aqueous ethanol (88% yield, mp 157–159°): homogeneous in system S6; uv max (CH₃OH) 236 m μ (ϵ 11,800); ir (KBr) 3510, 3385 (OH and NH), 1757 (ester C=O), 1706 (C-11 C=O), 1692 (amide C=O), 1573 (amide), and 1215 cm⁻¹ (ester COC). The dried product gained 4.1% in weight when exposed to the atmosphere. *Anal.* Calcd for C₃₅H₅₁O₁₃N₃: N, 5.82. Found: N, 5.60.

Methyl (21-Acetoxy-11,20-dioxo-5 β -pregnan-3 α -yl 2,3,4-Tri-O-acetyl- β -D-glucosid)uronate 20-Semicarbazone (23). A. From 19.—A solution of 750 mg of semicarbazide hydrochloride and 504 mg of NaHCO₃ in 2 ml of water was added to 695 mg of 20-oxo conjugate 19 in 20 ml of methanol. After 18 hr, crystals (709 mg, 93%, mp 178–181°) of chromatographically pure (system

(20) If the pH is adjusted to 3.0 and the solution is stirred for a few minutes, it remains homogeneous when the pH is subsequently brought to 2.1; under these conditions, addition of methanol is not necessary.

S6) semicarbazone were obtained from aqueous methanol: uv max (CH₃OH) 237 m μ (ϵ 12,800); ir (KBr) 3515, 3395, 3285 br (NH), 1753 (ester C=O), 1705 (C-11 + amide, C=O), 1570 (amide), and 1217 cm⁻¹ (ester COC). *Anal.* Calcd for C₃₇H₅₈O₁₄N₃·1/2H₂O: C, 57.50; H, 7.04; N, 5.44. Found: C, 57.55; H, 6.95; N, 5.38.

B. From 22 and 24.—Acetylation of 22 and 24 in 1:1 acetic anhydride-pyridine gave 23 (mp 178–181°).

Methyl (21-Hydroxy-11,20-dioxo-5 β -pregnan-3 α -yl β -D-Glucopyranosid)uronate 20-Semicarbazone (24). **A. From 20.**—Treatment of 50 mg of ester 20 with semicarbazide hydrochloride as described under the preparation of 23 from 19 gave crystals (30.8 mg, 55% yield) of 24 from butyl acetate.

B. From 23.—To a solution of 764 mg of semicarbazone 23 in a mixture of 10 ml of chloroform and 5 ml of methanol was added 5 ml of 0.04 N methanolic NaOH. After 45 min, a slight excess of 1.0 N acetic acid in methanol was added and the solution was taken to dryness immediately to remove all acetic acid and prevent hydrolysis of the semicarbazone. Hygroscopic crystals (552 mg, 92%, mp 173° dec) were obtained from methanol-butyl acetate: homogeneous in system S14; uv max (CH₃OH) 236 m μ (ϵ 11,800); ir (KBr) 3500–3300 (OH and NH), 1742 (ester C=O), 1702 sh (C-11 C=O), 1680 (amide C=O), and 1565 cm⁻¹ (amide). *Anal.* Calcd for C₂₉H₄₆O₁₀N₃·1/2H₂O: C, 57.59; H, 7.84; N, 6.95; CH₃O, 5.13. Found: C, 57.33; H, 7.64; N, 7.35; CH₃O, 5.53.

Methyl α -D-Glucopyranuronate Cyclic 1,2-(Hydrogen [S]-Orthoacetate) 3,4-Diacetate 3-Ester with 3 α ,21-Dihydroxy-5 β -pregnane-11,20-dione 21-Acetate (25).—The filtrate from the crystallization of 3.02 g of compound 19 (described under 19 from 3) was evaporated and the residue was chromatographed as described for the separation of 19 except that system S1 was used. The band at 3.1 HBV gave 1.64 g of residue which crystallized from methanol-water to yield 1.56 g (22%, mp 95–98°) of chromatographically pure (systems S1 and S4) orthoacetate 25: $[\alpha]_D + 78 \pm 2^\circ$ (CHCl₃); ir (KBr) 1753 (ester C=O), 1708 (ketone C=O), and 1225 cm⁻¹ (ester COC); nmr (CDCl₃) δ 0.61 (C-18 CH₃), 1.13 (C-19 CH₃), 1.75 (orthoacetate CH₃), 2.10 (3',4' OAc groups), 2.15 (C-21 OAc), 2.50 (C-12 protons), 3.78 (CH₃O of ester), 4.55 (C-21 methylene), and 5.81 ppm (doublet, 1-proton of glucuronyl group, $J_{1,2} = 5$ Hz). *Anal.* Calcd for C₃₈H₅₆O₁₄: C, 61.17; H, 7.13; CH₃O, 4.39; CH₃CO, 24.36. Found: C, 61.31; H, 7.36; CH₃O, 4.58; CH₃CO, 22.96.

21-Acetoxy-5 β -pregnane-3,11,20-trione.—Chromatography fractions 144–195 (HBV 2.2), described under 19 from 3, afforded crystals (462 mg, 12%, mp 157–160°) from methanol; this product was identified as 21-acetoxy-5 β -pregnane-3,11,20-trione¹⁷ by comparison with an authentic sample. Paper chromatography of compound 3 (starting material in this preparation) in system S4 showed that it contained no 21-acetoxy-5 β -pregnane-3,11,20-trione.

Compound 3 (781 mg, 2.0 mmol) was treated with 2.20 g (8.0 mmol) of freshly prepared silver carbonate in benzene as described for the preparation of 4 except that the methyl 2,3,4-tri-O-acetyl-1-bromo-1-deoxy- α -D-glucuronate was omitted. Chromatography of the product on 300 g of Celite (system S4) separated a band (HBV 2.0) which gave crystals (385 mg, 50%, mp 157–158°) of 21-acetoxy-5 β -pregnane-3,11,20-trione from aqueous methanol.

17 β -Carboxy-11-oxo-5 β -androstan-3 α -yl β -D-Glucopyranosiduronic Acid (26). **A. From 19.**—The residue (71 mg) from the band eluted at 1.14 HBV (described under 21 from 19) was dissolved in the mobile phase of system S13; crystals formed (7.5 mg, 3.0%, mp 214–216° dec). The mother liquor was chromatographed on 30 g of Celite in system S13. The band eluted at

7.5 HBV yielded crystals (12.5 mg, 5.0%, mp 214–215°) of 26 (from ethanol) identical with the sample prepared in the following paragraph.

B. From 27.—Compound 27 (325 mg) was hydrolyzed by treatment with alkali as described for the preparation of 21 from 23, procedure 1, and the solution was desalted on a column of Amberlite XAD-2. Crystals (228 mg, mp 214–215°) of chromatographically pure (system S13) 26 were obtained from ethanol: $[\alpha]_D + 43 \pm 2^\circ$ (CH₃OH); ir (KBr) 3380 (OH), 1763 (carboxyl C=O), 1710 sh, and 1696 cm⁻¹ (C-11 C=O). *Anal.* Calcd for C₂₈H₃₈O₁₀: C, 61.16; H, 7.50. Found: C, 61.18; H, 7.74.

Methyl (17 β -Carboxy-11-oxo-5 β -androstan-3 α -yl 2,3,4-Tri-O-acetyl- β -D-glucosid)uronate (27).—To 333 mg (0.50 mmol) of 18 in 15 ml of glacial acetic acid was added 1.54 mmol of H₅IO₆ in 70 ml of 80% acetic acid. After 15 min, water was added, the solution was extracted with ethyl acetate, and the extract was washed with water and taken to dryness. Crystals (230 mg, mp 220–222°; 61.5 mg, mp 218–220°) were obtained from ethanol. The sample for analysis melted partially at 135–140°, recrystallized spontaneously, and remelted at 220–221°: $[\alpha]_D + 41 \pm 2^\circ$ (CH₃OH); ir (KBr) 3320 (OH), 1756 (ester + carboxyl, C=O), 1705 (C-11 C=O), and 1212 cm⁻¹ (ester COC). *Anal.* Calcd for C₃₈H₄₆O₁₃·1/2H₂O: C, 60.08; H, 7.18; CH₃O, 4.70. Found: C, 60.35; H, 7.08; CH₃O, 4.83.

Methyl (17 β -Carbomethoxy-11-oxo-5 β -androstan-3 α -yl 2,3,4-Tri-O-acetyl- β -D-glucosid)uronate (28). **A. From 27.**—Treatment of 27 (100 mg) in methanol with diazomethane in ether gave ester 28 (101 mg, mp 172–173°), identical with the product prepared from 26.

B. From 26.—Treatment of 100 mg of a methanolic solution of 26 (derived from 27) with diazomethane in ether followed by acetylation with acetic anhydride-pyridine and crystallization from ethanol gave 121 mg (92%) of chromatographically pure (tlc) ester 28 (170–171°). The analytical sample had mp 172–173°; $[\alpha]_D + 36 \pm 2^\circ$ (CH₃OH); ir (KBr) 1770 sh, 1755 (acetate C=O), 1740 (ester C=O), 1705 (C-11 C=O), and 1210 cm⁻¹ (ester COC). *Anal.* Calcd for C₃₄H₄₈O₁₃·H₂O: C, 59.81; H, 7.38; CH₃O, 9.09. Found: C, 60.09; H, 7.16; CH₃O, 8.99.

When dicarboxylic acid 26 (derived from 18 with alkali) was esterified with diazomethane and acetylated, 28 was obtained.

Registry No.—1, 566-03-0; 2, 2631-05-2; 3, 2526-11-6; 4, 36707-52-5; 5, 36707-53-6; 6, 36707-54-7; 7, 36707-55-8; 8, 36707-56-9; 9, 36707-57-0; 10, 36763-74-3; 11, 36763-75-4; 12, 36707-58-1; 13, 36707-59-2; 14, 36707-60-5; 15, 36707-61-6; 16, 36707-62-7; 17, 36707-63-8; 18, 36707-05-8; 19, 36707-06-9; 20, 36707-07-0; 21, 35105-23-8; 22, 36707-09-2; 23, 36707-10-5; 24, 36707-11-6; 25, 36707-12-7; 26, 36707-13-8; 27, 36707-14-9; 28, 36707-15-0; 21-acetoxy-5 β -pregnane-3,11,20-trione, 36707-16-1.

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